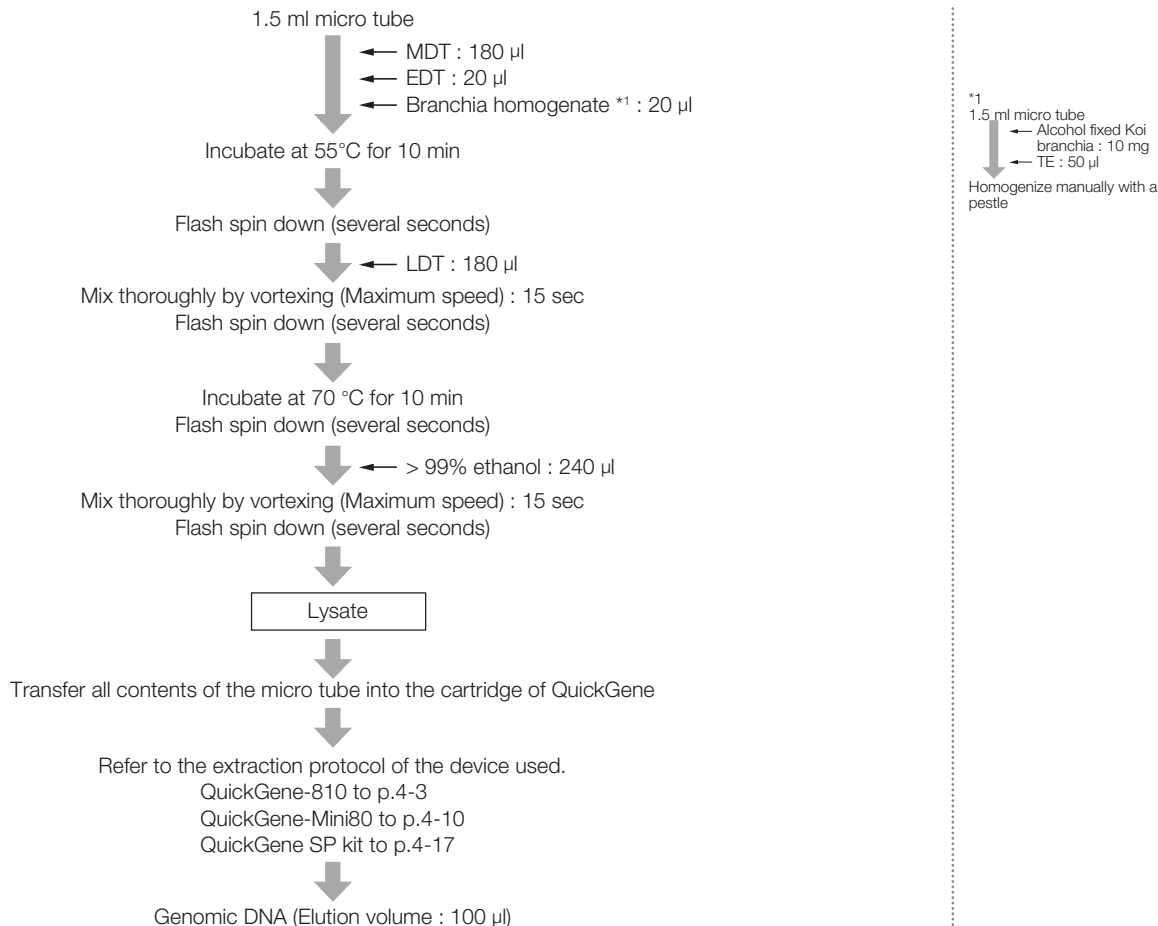


Chapter 3-IX

Genomic DNA Extraction from Virus

Genomic DNA Extraction from Branchia of Koi Herpes Virus (KHV) Infected Fish

Protocol



Results

Electropherogram

No Data

The yield of genomic DNA

	No.	Yield (µg)
Normal fish	1	4.24
	2	4.07
Infected fish	1	0.67
	2	1.28
	3	2.41
	4	2.35

Protein contamination : A260/280

	No.	A/260/280
Normal fish	1	2.19
	2	2.27
Infected fish	1	2.04
	2	2.39
	3	2.10
	4	1.99

Chaotropic salt contamination : A260/230

No Data

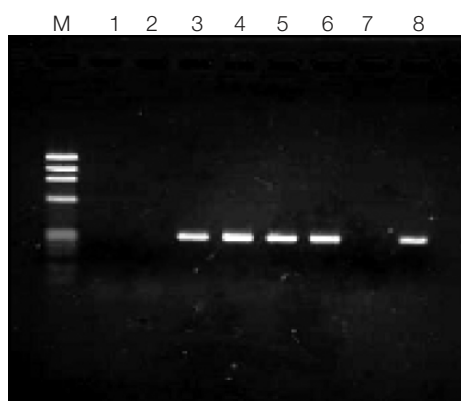
Other

• PCR

DNA isolated by using QuickGene-810 system was used for PCR template.

PCR was performed according to the method by Yuasa et al, Improvement of a PCR method with the *Sph* 1-5 primer set for the detection of Koi herpesvirus (KHV), Fish Pathology, 40, 37-39 (2005).

Primer : *Sph* I -5F, *Sph* I -5R



M : λ x 174-*Hae* III digest

1 : Normal fish No.1

2 : Normal fish No.2

3 : Infected fish No.1

4 : Infected fish No.2

5 : Infected fish No.3

6 : Infected fish No.4

7 : Negative control

8 : Positive control

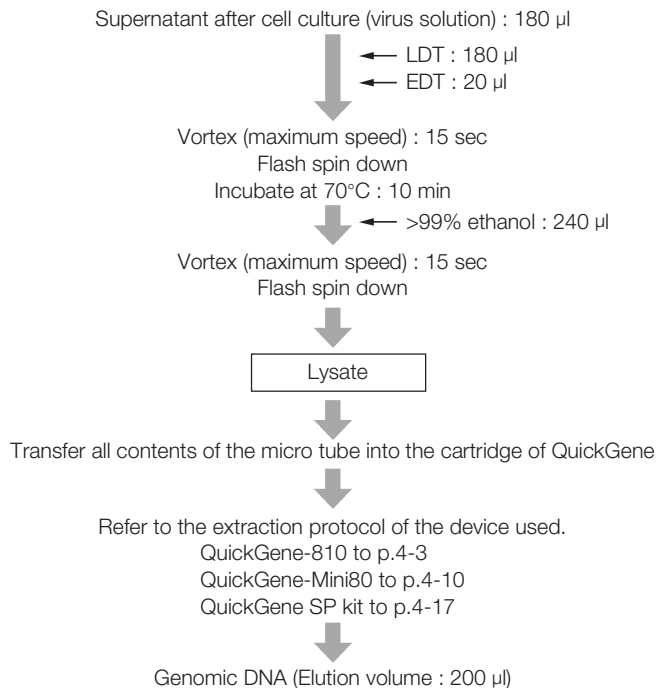
PCR amplification similar to that for positive control was confirmed for infected fish, No.1-4.

Common protocol is usable for the following

No Data

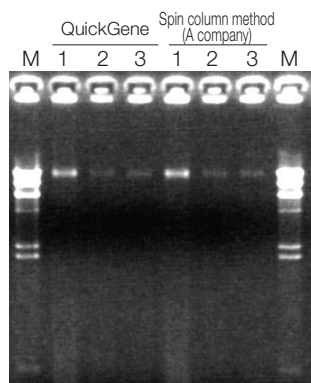
Genomic DNA Extraction from Herpes Simplex Virus-type 1 (HSV-1) Virus Solution

Protocol



Results

Electropherogram



Electrophoresis condition : 1.5% agarose / 1 x TAE

M : λ -Hind III
 1 : No.1 VR3 (wild strain)
 2 : No.2 d41 (UL41 defective mutant)
 3 : No.3 d13 (UL13 defective mutant)

No decomposition was detected for extracted genomic DNA.

The yield of genomic DNA

sample	No.1	No.2	No.3
QuickGene	324 ng	32 ng	51 ng
Spin column method (A company)	351 ng	36 ng	40 ng

Protein contamination : A260/280

sample	No.1	No.2	No.3
QuickGene	2.23	2.01	2.14
Spin column method (A company)	1.98	2.41	1.92

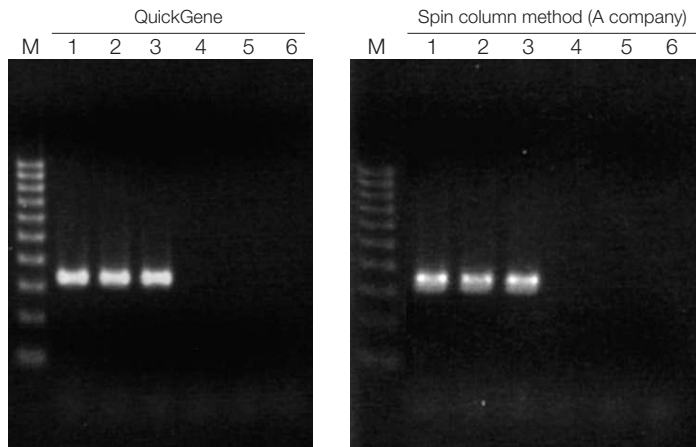
Chaotropic salt contamination : A260/230

No Data

Other

• PCR

HSV-1 gene was detected by PCR with HSV-1 specific primer and HSV-2 specific primer for genomic DNA extracted from HSV-1 using QuickGene system and Spin column method (A company).



Electrophoresis condition : 2% agarose / 1 x TAE

M : 100 bp DNA Ladder
1 : No.1 VR3/HSV-1 primer
2 : No.2 d41/HSV-1 primer
3 : No.3 d13/HSV-1 primer
4 : No.1 VR3/HSV-2 primer
5 : No.2 d41/HSV-2 primer
6 : No.3 d13/HSV-2 primer

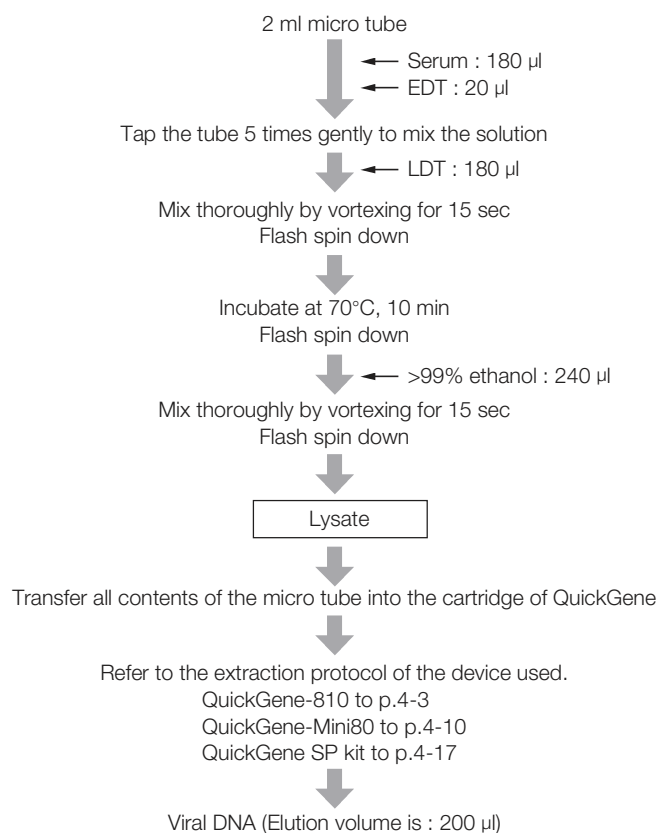
PCR products were detected for each genomic DNA.

Common protocol is usable for the following

No Data

HBV DNA Extraction from Serum

Protocol



Results

■ Electropherogram

No Data

■ The yield of viral DNA

No Data

■ Protein contamination : A260/280

No Data

■ Chaotropic salt contamination : A260/230

No Data

■ Other

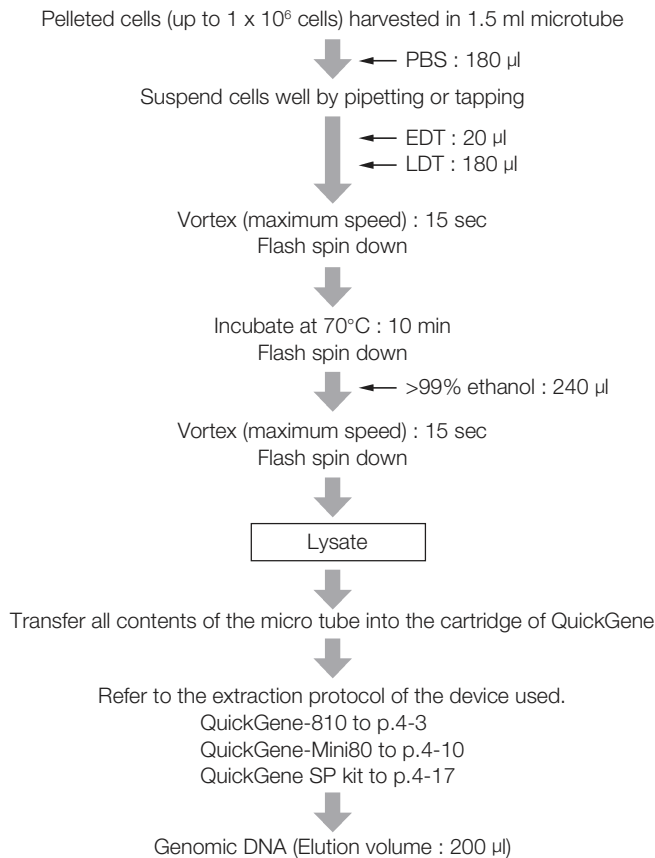
No Data

Common protocol is usable for the following

No Data

Human Papiloma Virus (HPV) DNA Extraction from Human Cervical Carcinoma Cell lines

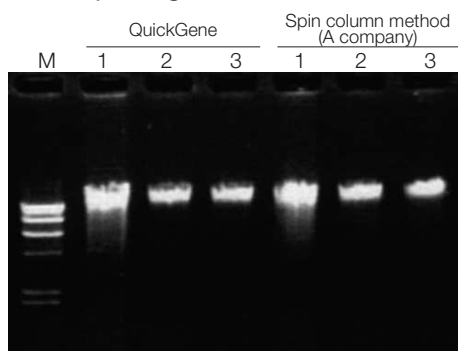
Protocol



Results

Cell strain : HeLa (containing 10 ~ 50 copies of HPV18)
: SiHa (containing 1 ~ 2 copies of HPV16)
: Caski (containing 400 ~ 600 copies of HPV16)

Electropherogram



Electrophoresis condition : 1.5% agarose / 1 x TAE

M : λ -Hind III
1 : HeLa
2 : SiHa
3 : Caski

No decomposition was detected for extracted genomic DNA.

The yield of genomic DNA

sample	HeLa	SiHa	Caski
QuickGene	23.5 µg	11.6 µg	13.5 µg
Spin column method (A company)	26.2 µg	10.5 µg	7.3 µg

Protein contamination : A260/280

sample	HeLa	SiHa	Caski
QuickGene	2.00	1.94	1.93
Spin column method (A company)	1.81	1.94	2.15

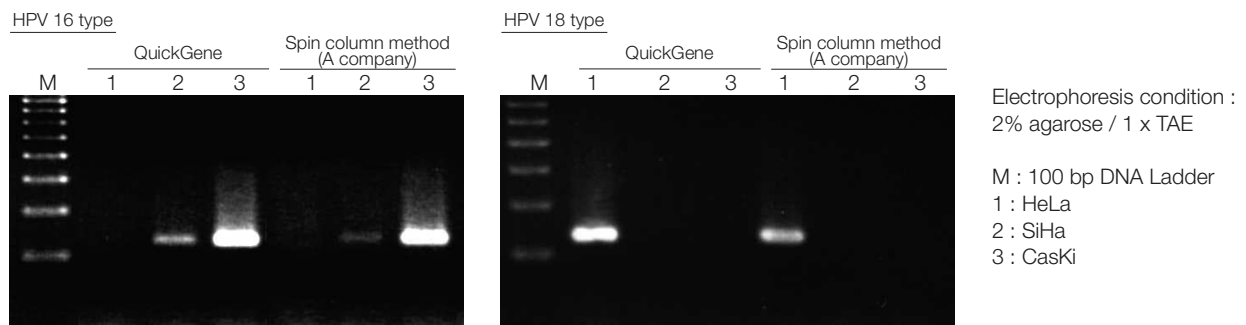
Chaotropic salt contamination : A260/230

No Data

Other

• PCR

Viral genomic DNA of HPV 16 type and HPV 18 type was detected by PCR for genomic DNA extracted using QuickGene system and Spin column method (A company).



1 to 2 copies of HPV genomic DNA were detected per cell by PCR for HPV DNA extracted using QuickGene system.

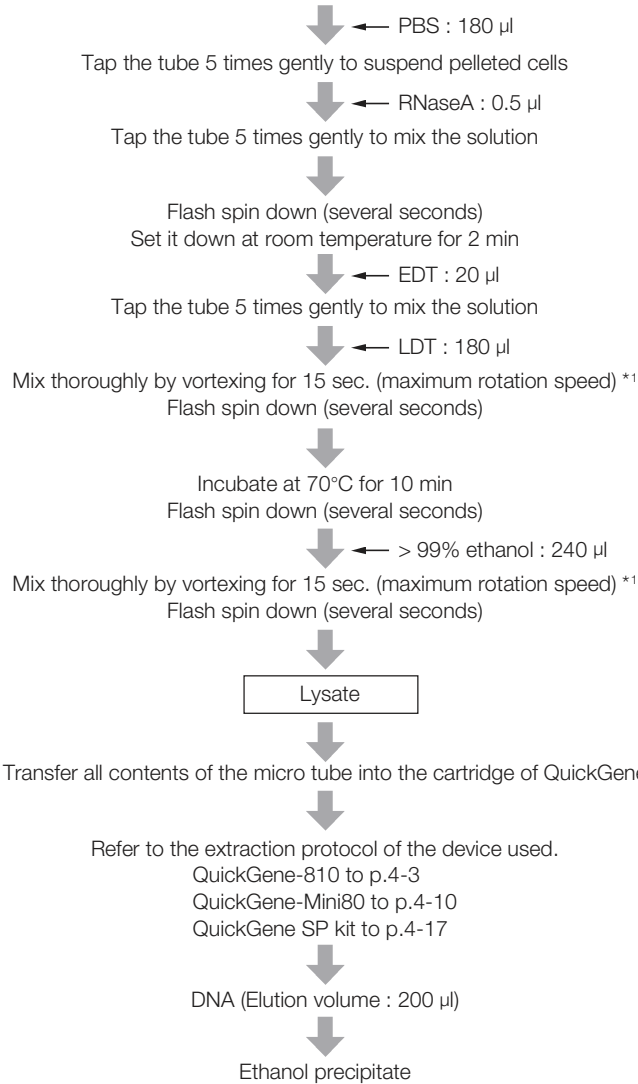
Common protocol is usable for the following

No Data

Viral DNA Extraction from Simian Immunodeficiency Virus (SIV) Infected Cells

Protocol

Place cells into 1.5 ml micro tube and pelletize (-1×10^6 cells in 1.5 ml micro tube)



*1 Mix completely by vortexing at the maximum speed. If the mixing is not enough by vortexing, use tapping, pipetting or inverting.

Results

■ Electropherogram

No Data

■ The yield of viral DNA (µg)

Time after infection (h)	1.5		3		6		24	
Virus	mock	SIV	mock	SIV	mock	SIV	mock	SIV
Cell number	1×10^6	1×10^6	1×10^6	8×10^5	1×10^6	9.2×10^5	1×10^6	1×10^6
QuickGene-810	7.6	7.9	3.0	8.0	4.5	8.0	8.2	7.4
Spin column	3.8	4.3	3.0	2.5	5.4	5.5	4.7	3.4

Protein contamination : A260/280

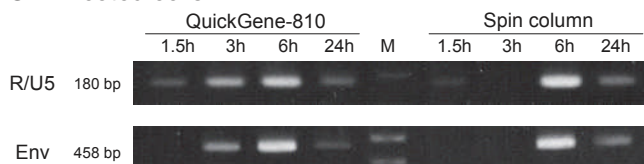
Time after infection (h)	1.5		3		6		24	
Virus	mock	SIV	mock	SIV	mock	SIV	mock	SIV
QuickGene-810	1.81	1.80	1.79	1.75	1.80	1.80	1.80	1.82
Spin column	1.85	1.85	1.8	1.81	1.79	1.77	1.81	1.82

Chaotropic salt contamination : A260/230

No Data

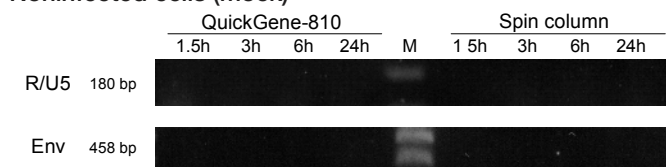
Other

- AGE of PCR fragments of DNA

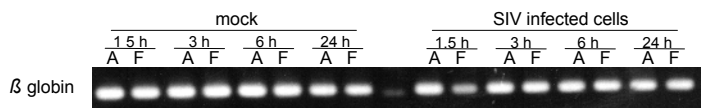
SIV infected cells


PCR was performed with 1 mg of DNA isolated from SIV-infected cells using the QuickGene-810 system and spin column.

The electrophoretic band of PCR amplified products of DNA isolated 1.5 hours and 3 hours after infection by using QuickGene-810 system could be detected.

Noninfected cells (mock)


M : marker(ladder)



F : QuickGene-810
A : Spin column

Common protocol is usable for the following

No Data



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